

5

TECHNICAL FIELD

10

050601-0984980

15

30

35

transcript RNA for CHIP28, the current designation of which is AQP1, had been introduced showed increased water permeability. In Science (vol. 264, pp. 92 to 95, 1994), B. A. van Oost et al. disclosed the amino acid sequence of human AQP2 and suggested that this should be involved in vasopressin-dependent urine concentration.

In Proc. Natl. Acad. Sci. USA (vol. 91, pp. 6269 to 6273, 1994), Ishibashi et al. disclosed the nucleotide sequence of the gene for renal collecting tubule-derived AQP3 and the amino acid sequence encoded thereof. Ishibashi et al. confirmed its water channel activity by injecting the AQP3 cRNA into Xenopus laevis oocytes and measuring the water permeability thereof. Ishibashi et al. reported that this AQP3 transported not only water but also nonionic small molecules such as urea and glycerol.

In Proc. Natl. Acad. Sci. USA (vol. 91, pp. 13052 to 13056, 1994), J. S. Jung et al. reported about the isolation of AQP4. This AQP4 is known to occur most abundantly in mammalian brains and have mercury resistance. In J. Biol. Chem. (vol. 270, pp. 1908 to 1912, 1995), S. Raina et al. who prepared rat salivary gland-derived AQP5 cDNA describe the nucleotide sequence of the cDNA and the amino acid sequence encoded thereby. S. Raina et al. cloned the cDNA by utilizing the occurrence of an NPA sequence and confirmed its function by observing that the cRNA enhances the water permeability of Xenopus laevis oocytes.

The aquaporin family mentioned above is considered to be involved in water metabolism in mammals and, for example, it has been confirmed that AQP2 is found only in the renal collecting tubule luminal membrane, which is indicative of its close association with the vasopressin-urea concentration system, and its involvement in renal diseases has become acknowledged. Therefore, such membrane proteins having water channel activity are of importance in any attempt to develop novel therapies for water-associated diseases.

Meanwhile, the expression of the aquaporin family

09849980-050801

mentioned above has been confirmed in such organs as kidney, brain, gall bladder, eye, intestine, salivary gland and bronchus but there is no report as yet about the occurrence of membrane proteins having water channel activity in other organs or tissues, particularly in adipose tissue.

SUMMARY OF THE INVENTION

In view of the above-mentioned state of the art, the present invention has for its object to provide a novel membrane protein having water channel activity and a DNA sequence encoding for the polypeptide.

The present invention is related to a novel polypeptide having water channel activity which has the amino acid sequence, within the molecule thereof, shown in the sequence listing under SEQ ID NO:1.

The present invention is also related to a nucleotide sequence itself which codes for a polypeptide having, within the molecule thereof, the amino acid sequence shown in the sequence listing under SEQ ID NO:1 and having water channel activity.

The present invention is further related to the DNA sequence shown in the sequence listing under SEQ ID NO:2.

The present invention is still further related to a polypeptide having water channel activity which has the amino acid sequence, within the molecule thereof, encoded by the nucleotide No. 173 to No. 1198 of the nucleotide sequence shown in the sequence listing under SEQ ID NO:2.

DETAILED DESCRIPTION OF THE INVENTION

In the following, the present invention is described in detail.

The polypeptide of the present invention has the amino acid sequence shown in the sequence listing under SEQ ID NO:1. This polypeptide has a sequence composed of three amino acids, namely asparagine-proline-alanine, as the amino acid Nos. 195

09849980-050801

to 197. However, the characteristic feature common to the so-far known AQPs, that said asparagine-proline-alanine sequence occurs twice, is not found in the polypeptide of the present invention. That this polypeptide has water channel activity can be confirmed from the fact that it enhances the water permeability of *Xenopus laevis* oocytes.

The above polypeptide may be generated by translation by a protein synthesis system constituted, in vivo or in vitro, based on the nucleotide sequence coding for the amino acid sequence of said polypeptide. The nucleotide sequence of the present invention substantially has a region coding for the amino acid sequence of said polypeptide and, where necessary, may contain one or more other regions such as a promoter region. In the protein synthesis based on genetic information, the information carried by the gene DNA is transcribed into mRNA as the result of DNA-dependent RNA synthesis aided by RNA polymerase. And, this mRNA is translated into the amino acid sequence in a tRNA-containing protein synthesis system. Therefore, the nucleotide sequence of the present invention includes not only the DNA sequence but also the RNA sequence. Furthermore, since it is generally known that, for an amino acid, there is one or a plurality of codons corresponding thereto, it is a matter of course that the above-mentioned nucleotide sequence is not limited to only one sequence but may include nucleotide sequences resulting from substitution of another synonymous codon coding for the same amino acid.

The above polypeptide can be formed based on the genetic information carried by the DNA sequence shown in the sequence listing under SEQ ID NO:2. This polypeptide is encoded by that portion of the nucleotide sequence shown in the sequence listing under SEQ ID NO:2 which ranges from the nucleotide No. 173 to No. 1198. Of the DNA sequence shown in the sequence listing under SEQ ID NO:2, the nucleotide sequences other than the portion of said nucleotide numbers are noncoding regions, among which the polyadenylation consensus sequence AATAAA occurs at

the nucleotide No.1234 to No.1239. Other possible reading frames of said DNA sequence shown under SEQ ID NO:2 can be excluded from consideration, since the polypeptides encoded are very small-sized, hence considered to be incapable of performing any water channel function.

It has been confirmed by the inventors that the full-length sequence of the above nucleic acid bases has no counterpart sequence either in GenBank or in dbEST.

The polypeptide of the present invention has water channel activity in adipose tissue. While adipose tissue is distributed in various parts of the living organism, the polypeptide of the present invention has an action to control the transfer of water in such adipose tissue and is expected to be effective in upholding normal functions of adipose tissue at various sites.

BEST MODES FOR CARRYING OUT THE INVENTION

The above-mentioned DNA sequence given under SEQ ID NO:2 corresponds to the nucleotide sequence of cDNA obtained from human adipose tissue by cloning. Human adipose tissue is a tissue which stores fat as energy reserves. It is known that various proteins are formed in this adipose tissue. A 3'-directed DNA library is known as a cDNA library from which the genes actually expressed in this adipose tissue or, in other words, the mRNA composition in this adipose tissue can be copied faithfully. This 3'-directed DNA library contains only those specified 3'-terminal regions of mRNAs which range from poly(A) to the MboI site which is a restriction enzyme recognition site upstream of said poly(A) and, therefore, said library is suited for template preparation by the PCR technique. Therefore, by extracting a clone from this library and using it to determine a longer nucleotide sequence including the amino acid coding region from this complete adipose tissue cDNA library, it becomes possible to obtain the genetic information concerning the protein which is actually formed in adipose tissue. The

0984980-050801

DNA sequence of the present invention as shown under the above-mentioned SEQ ID NO:2 is found by such cloning. A method of obtaining the cDNA by cloning from human adipose tissue is now described in detail.

5 Known as said method is, for example, the method described in Biochem. Biophys. Res. Commun., 221, 286 to 289 (1996). According to this method, the total RNA is first separated from adipose tissue and, when necessary, purified to give poly(A) RNA. For this purification, commercially available
10 purification kits can be used. For example, Pharmacia's Quick prep mRNA purification kit or the like in which oligo(dT)-cellulose and various buffers are used in combination can judiciously be employed. Then, a double-stranded cDNA is synthesized using a pUC19 system vector primer and the
15 double-stranded cDNA so synthesized is selectively cleaved with the restriction enzyme MboI (which recognizes the nucleotide sequence GATC). On that occasion, the GATC sequence on the vector molecule side, which can be methylated to give G^mATC when replication is effected in dam⁺ bacterial cells, is not cleaved
20 with MboI. As the cleaved cDNA is subjected to self-cyclization using E. coli ligase, a plasmid containing a cDNA fragment extending from poly(A) to the nearest MboI site is completed. This plasmid is introduced into Escherichia coli, followed by cultivation and selection of a transformant E. coli colony.
25 Then, the cDNA in said colony is amplified by the PCR technique using appropriate PCR primers.

On the other hand, the full-length double-stranded cDNA synthesized using the pUC19 system vector primer is cleaved at the 5' end using T4 polymerase and subjected to cyclization
30 using T4 ligase and introduction into Escherichia coli for transformation. From among the thus-obtained transformant colonies, the desired colony is obtained by screening using, as a probe, a labeled form of the adipose tissue-specific cDNA obtained from said 3'-directed DNA library by the method
35 mentioned above. The insert cDNA in this colony is amplified

0984980-050801

The nucleotide sequence of the thus-cloned cDNA can be determined, for example, by reaction with a primer dye, purification and analysis using an automated sequencer or the like. In this manner, the DNA sequence of the present invention can be obtained.

The polypeptide of the present invention has water channel activity. This water channel activity can be confirmed by observing an enhancement of water permeability in Xenopus laevis oocytes. It is known that no AQP family gene has been expressed in Xenopus laevis oocytes and, therefore, any increase in water permeability as caused by the injected mRNA can be easily confirmed. For this reason, said oocytes are widely used in confirming water channel activity. For example, in Proc. Natl. Acad. Sci. USA, vol. 91, pp. 6269 to 6273 (1994), Ishibashi et al. confirmed the water channel activity by inserting the AQP3 cDNA into the pSP64T-derived BlueScript vector, synthesizing the cRNA using T7 RNA polymerase, injecting this cRNA into Xenopus laevis oocytes and, after 48 to 62 hours of incubation following injection, observing an increase in water permeability and in the volume of the oocyte.

The following examples illustrate the present invention in further detail. These examples are, however, by no means
35 limitative of the scope of the present invention.

Example 1

Determination of DNA nucleotide sequence

According to a recent report [Biochem. Biophys. Res. Commun., 221, 286 to 289 (1996)], a DNA for an adipose tissue-specific collagen-like factor has been cloned using a 3'-directed DNA library containing only those specified 3'-terminal regions of RNA molecules which range from poly(A) to the restriction enzyme MboI site upstream thereof [Nature Genet., 2, 173 to 179 (1992)], which library makes it possible not only to identify the gene under expression but also to examine the frequency of expression or amount of expression. In accordance with the method described in the above report, the DNA nucleotide sequence of the present invention was determined using an adipose tissue-specific 3'-directed DNA library.

Method

Reverse transcriptase was added to poly(A)⁺ RNA separated and purified from human adipose tissue using a Quick prep mRNA purification kit, followed by insertion into λ ZAP II containing the pUC19 system vector pBluescript and introduction into Escherichia coli. Screening carried out using an adipose tissue-specific partial DNA as a probe gave a colony of transformant Escherichia coli. Then, together with two primers (SK: 5'CGCTCTAGAACTAGTGGATC3'; T7: 5'GTAATACGACTCACTATAGGGC3'), PCR [polymerase chain reaction; (30 seconds at 95°C + 30 seconds at 50°C + 60 seconds at 70°C) x 15 cycles, followed by (30 seconds at 95°C + 60 seconds at 70°C) x 15 cycles], was carried out and, after sonication, the product was subcloned in M13. A primer dye was added thereto and, after purification, the nucleotide sequence of the DNA was analyzed using an automated sequencer. The nucleotide sequence obtained is shown in the sequence listing under SEQ ID NO:2.

Example 2

Examination as to water permeability

As for the studies on the water permeability of the AQP family, there are reports about AQP1 [Science, 256, 385-387 (1992)] and AQP3 [Proc. Natl. Acad. Sci. USA, 91, 6269-6273 (1994)] in which RNA was introduced into *Xenopus laevis* oocytes, in which AQP family genes have not been expressed, and then the water permeability was calculated from changes in the surface area and volume of said oocytes in a hypotonic culture medium. Therefore, the membrane protein encoded by the DNA of the present invention was checked for water permeability according to the method described in the references cited above.

Test method

The RNA (10 ng) obtained in Example 1 from human adipose tissue was introduced into *Xenopus laevis* oocytes by microinjection and the oocytes were incubated at 20°C for 3 days in an isotonic culture medium (about 200 mOsm). The cultured oocytes were transferred to a hypotonic culture medium (about 40 mOsm). Photograph was taken 20 seconds and 40 seconds after transfer, and the sectional area and volume of each oocyte were determined using an image analyzer. The water permeability of the membrane protein was calculated as follows:

$$\text{Permeability (cm/sec)} = [(V_{40} - V_{20})/20] / [(A_{20} \times 10^{-2} \times 4) \times 1.384]$$

where V_{20} denotes the oocyte volume (cm^3) after 20 seconds, V_{40} denotes the oocyte volume (cm^3) after 40 seconds, and A_{20} denotes the oocyte sectional area (mm^2) after 20 seconds.

The result is shown in Table 1. The water permeability found when purified water was used for microinjection in lieu of the RNA is also shown.

Further, the expression of the polypeptide of the present

000050 08664860

invention in said oocytes was confirmed by a C-terminal region immunoassay using the rabbit antiserum obtained by using the polypeptide of the present invention as synthesized in vitro.

5 Table 1

	Water permeability (cm/sec)
Group in which RNA introduction was not made	30.0×10^{-4}
Group in which RNA introduction was made	292.5×10^{-4}

As is evident from Table 1, the polypeptide encoded by the DNA sequence of the present invention caused an increase in water permeability after introduction into the oocytes.

10 This result indicate that the polypeptide of the present invention has water channel activity.

INDUSTRIAL UTILIZABILITY

15 The present invention provides a novel protein having water channel activity and a novel DNA sequence encoding the protein. Said protein is one found in human adipose tissue for which the occurrence of water channels has not been reported as yet. The present invention makes it possible to develop novel therapies for water or fat metabolism-associated diseases
20 in which said tissue is involved.

09849930-050801

[Sequence listings]

SEQ ID No.: 1

Length: 342

5 Type: amino acid

Topology: linear

Species: peptide

Sequence

10 Met Val Gln Ala Ser Gly His Arg Arg Ser Thr Arg Gly Ser Lys Met
 5 10 15
 Val Ser Trp Ser Val Ile Ala Lys Ile Gln Glu Ile Leu Gln Arg Lys
 20 25 30
 Met Val Arg Glu Phe Leu Ala Glu Phe Met Ser Thr Tyr Val Met Met
 15 35 40 45
 Val Phe Gly Leu Gly Ser Val Ala His Met Val Leu Asn Lys Lys Tyr
 50 55 60
 Gly Ser Tyr Leu Gly Val Asn Leu Gly Phe Gly Phe Gly Val Thr Met
 65 70 75 80
 20 Gly Val His Val Ala Gly Arg Ile Ser Gly Ala His Met Asn Ala Ala
 85 90 95
 Val Thr Phe Ala Asn Cys Ala Leu Gly Arg Val Pro Trp Arg Lys Phe
 100 105 110
 Pro Val Tyr Val Leu Gly Gln Phe Leu Gly Ser Phe Leu Ala Ala Ala
 25 115 120 125
 Thr Ile Tyr Ser Leu Phe Tyr Thr Ala Ile Leu His Phe Ser Gly Gly
 130 135 140
 Gln Leu Met Val Thr Gly Pro Val Ala Thr Ala Gly Ile Phe Ala Thr
 145 150 155 160
 30 Tyr Leu Pro Asp His Met Thr Leu Trp Arg Gly Phe Leu Asn Glu Ala
 165 170 175
 Trp Leu Thr Gly Met Leu Gln Leu Cys Leu Phe Ala Thr Thr Asp Gln
 180 185 190
 Glu Asn Asn Pro Ala Leu Pro Gly Thr Glu Ala Leu Val Ile Gly Ile
 35 195 200 205

09849980-050801

Leu Val Val Ile Ile Gly Val Ser Leu Gly Met Asn Thr Gly Tyr Ala
 210 215 220
 Ile Asn Pro Ser Arg Asp Leu Pro Pro Arg Ile Phe Thr Phe Ile Ala
 225 230 235 240
 5 Gly Trp Gly Lys Gln Val Phe Ser Asn Gly Glu Asn Trp Trp Trp Val
 245 250 255
 Pro Val Val Ala Pro Leu Leu Gly Ala Tyr Leu Gly Gly Ile Ile Tyr
 260 265 270
 Leu Val Phe Ile Gly Ser Thr Ile Pro Arg Glu Pro Leu Lys Leu Glu
 10 275 280 285
 Asp Ser Val Ala Tyr Glu Asp His Gly Ile Thr Val Leu Pro Lys Met
 290 295 300
 Gly Ser His Glu Pro Thr Ile Ser Pro Leu Thr Pro Val Ser Val Ser
 305 310 315 320
 15 Pro Ala Asn Arg Ser Ser Val His Pro Ala Pro Pro Leu His Glu Ser
 325 330 335
 Met Ala Leu Glu His Phe
 340

T08050-08664860

SEQ ID No: 2

Length: 1258

Type: nucleotide

Strandedness: double strand

5 Topology: linear

Species: cDNA to mRNA

Original source

Organism: human

Tissue type: adipose tissue

10 Feature Information

Name/Key: peptide

Location: 173..1198

Identification method: E

15 Sequence

GGCTCTGGAC	TGGGGACACA	GGGATAGCTG	AGCCCCAGCT	GGGGGTGGAA	GCTGAGCCAG	60
GGACAGTCAC	GGAGGAACAA	GATCAAGATG	CGCTGTAACT	GAGAAGCCCC	CAAGGCGGAG	120
GCTGAGAATC	AGAGACATTT	CAGCAGACAT	CTACAAATCT	GAAAGACAAA	AC ATG GTT	178
					Met Val	
					1	
CAA GCA TCC GGG CAC AGG CGG TCC ACC CGT GGC TCC AAA ATG GTC TCC						226
Gln Ala Ser Gly His Arg Arg Ser Thr Arg Gly Ser Lys Met Val Ser						
5 10 15						
TGG TCC GTG ATA GCA AAG ATC CAG GAA ATA CTG CAG AGG AAG ATG GTG						274
Trp Ser Val Ile Ala Lys Ile Gln Glu Ile Leu Gln Arg Lys Met Val						
20 25 30						
CGA GAG TTC CTG GCC GAG TTC ATG AGC ACA TAT GTC ATG ATG GTA TTC						322
Arg Glu Phe Leu Ala Glu Phe Met Ser Thr Tyr Val Met Met Val Phe						
35 40 45 50						
GGC CTT GGT TCC GTG GCC CAT ATG GTT CTA AAT AAA AAA TAT GGG AGC						370
Gly Leu Gly Ser Val Ala His Met Val Leu Asn Lys Lys Tyr Gly Ser						
55 60 65						
TAC CTT GGT GTC AAC TTG GGT TTT GGC TTC GGA GTC ACC ATG GGA GTG						418
Tyr Leu Gly Val Asn Leu Gly Phe Gly Phe Gly Val Thr Met Gly Val						
35 70 75 80						

0984980 050801

T-08090-0864860

TTC ATT GGC TCC ACC ATC CCA CGG GAG CCC CTG AAA TTG GAG GAT TCT 1042
 Phe Ile Gly Ser Thr Ile Pro Arg Glu Pro Leu Lys Leu Glu Asp Ser
 275 280 285 290
 GTG GCG TAT GAA GAC CAC GGG ATA ACC GTA TTG CCC AAG ATG GGA TCT 1090
 5 Val Ala Tyr Glu Asp His Gly Ile Thr Val Leu Pro Lys Met Gly Ser
 295 300 305
 CAT GAA CCC ACG ATC TCT CCC CTC ACC CCC GTC TCT GTG AGC CCT GCC 1138
 His Glu Pro Thr Ile Ser Pro Leu Thr Pro Val Ser Val Ser Pro Ala
 310 315 320
 10 AAC AGA TCT TCA GTC CAC CCT GCC CCA CCC TTA CAT GAA TCC ATG GCC 1186
 Asn Arg Ser Ser Val His Pro Ala Pro Pro Leu His Glu Ser Met Ala
 325 330 335
 CTA GAG CAC TTC TAAGCAGAGA TTATTTGTGA TCCCATCCAT TCCCCAATAA 1238
 Leu Glu His Phe
 15 340
 AGCAAGGCTT GTCCGACAAA 1258

T08050-08664860